

A CRITICAL ASSESSMENT OF CURRENT VITRIFICATION METHODS

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Disclosure

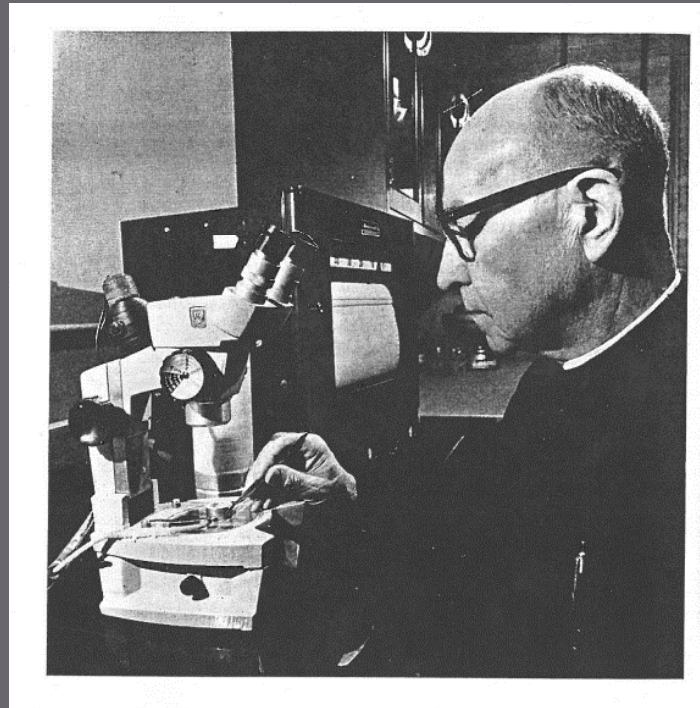
Scientific Director for The World Egg Bank
A For-Profit Company providing Oocyte
Donors and Cryopreserved Donor oocytes to
Fertility Clinics and Patients in the US and
Worldwide

No financial interests in any products being
Marketed to IVF Clinics other than TWEB
Products and Services

Goals:

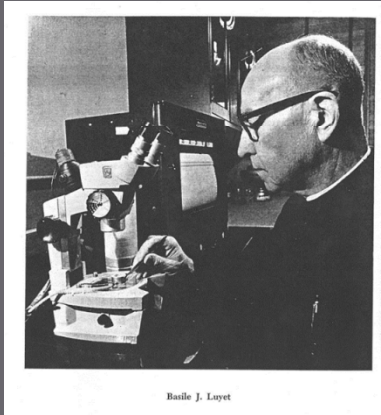
- ❑ Learn about the historical developments of Ultra-Rapid Vitrification
- ❑ Understand the sensitivities of current vitrification methods
- ❑ Explain what we should know, and yet do not
- ❑ Interpret the sensitivities and use that information as a guide to day-to-day practices
- ❑ Be able to examine a vitrification protocol and determine if the method is likely to be robust/repeatable

History of vitrification by “ultra-rapid” cooling



Basile J. Luyet (1897 – 1974)

Photo from: Meryman HT. Basile J. Luyet: In Memoriam. Cryobiology 1975;12:283-92.



Luyet. 1940

Life and Death at Low Temperatures

“One will obtain rapid elimination of heat by reducing the material to sheets with the smallest possible thickness and the largest possible area”. P. 208

“In order to reduce the heat capacity of the preparation, one must use very thin supports.” P. 218

“Instead of thin supports, we sometimes used, with advantage, a ring of about two millimeters in diameter, made of as thin a metal wire as possible and fastened to a light, rigid rod. One simply dips this loop into the culture and thus obtains, in the thin film within the ring, quite a considerable number of organisms.” P. 218

Photo from: Meryman HT. Basile J. Luyet: In Memoriam. Cryobiology 1975;12:283-92.
Text from Luyet BJ, Gehenio PM. Life and death at low temperatures. Normandy, Missouri: Biodynamica, 1940;341.

representing a ${}^1\Sigma - {}^1\Sigma$ transition, gave $B_1' = 0.5747$ and $B_2' = 0.5797 \text{ cm.}^{-1}$. The agreement is good.

Our analysis shows that the red system represents a ${}^1\Sigma - {}^4\Pi$ transition. A more detailed paper on the red bands will appear in *Arkiv för Fysik*.

ALBIN LAGERQVIST
ULLA UHLER

Physics Department,
University of Stockholm,
April 23.

¹ Mahard, P. C., *Phys. Rev.*, **42**, 609 (1932).

² Mahard, P. C., *Ind. J. Phys.*, **9**, 455 (1933).

³ Lagerqvist, A., *Ark. f. Mat., Astr. o. Fys.*, **29 A**, No. 25 (1943).

Revival of Spermatozoa after Vitrification and Dehydration at Low Temperatures

THE effect on spermatozoa of vitrification at temperatures of -79°C . and below has been studied by several authors. Human spermatozoa appear to be the most resistant; a substantial proportion may show good motility on thawing after even prolonged vitrification. Revival is far better when semen is frozen in bulk than when minimal amounts in capillary tubes are used¹. No explanation of this result is yet forthcoming, but it would appear that rapidity of freezing is less important than the avoidance of surface effects.

Positive results have also been obtained with frog and fowl spermatozoa^{2,3}, though in both these cases

ing 40 per cent glycerol, the spermatozoa resume full motility on thawing. So far as retention of motility is concerned, the specimen is indistinguishable from its unvitrified control; it shows even the wave motion characteristic of fowl semen. Decreasing the final concentration of glycerol below 10 per cent decreases the protection against vitrification. Increasing it above 20 per cent results in progressive immobilization of the spermatozoa, which cannot altogether be reversed by further dilution with Ringer's solution; but with these higher concentrations, no additional loss of motility is caused by vitrification. Specimens of spermatozoa have been found to resume motility completely after long periods (up to ten weeks) of vitrification. Other experiments showed that both propylene glycol and ethylene glycol were more toxic than glycerol, and in relation to their toxicity less protective against vitrification.

The fact that spermatozoa resumed full motility after vitrification under the conditions described above made it possible to investigate the effects of freeze-drying. 1 c.c. of fowl semen was diluted with 1 c.c. of 20 per cent glycerol in Ringer's solution, and vitrified as a thin layer in a 100-c.c. distilling flask at -79°C . The temperature was then allowed to rise to -25°C . and the flask connected to a high-vacuum distillation system of which the condenser unit contained liquid air. After 3 hr., when the distillation was stopped, the semen had the appearance of being dry, and 1.7 c.c. of water was thawed from the condenser. While still cold, the dehydrated semen was

Ribosenucl

Is a recent nuclei of erythrocytes and ribonucleic acid a found that they contain a small ribosenucleic interest to be present in the

The erythrocyte method previously described by Thannhauser and Thannhauser determined the colour reaction of ribonucleic acid. The filtrate, nucleic acid, ribosenucleic acid, as determined by Neijbaum⁴, as quartz spectra values expressed in terms of nucleic acid are shown

Name of the s



1990s

Steponkus PL, Myers SP, Lynch DV, Gardner L, Bronshteyn V, Leibo SP, Rall WF, Pitt RE, Lin TT, MacIntyre RJ. Cryopreservation of *Drosophila melanogaster* embryos. *Nature* 1990;345:170-2.

Bovine Blastocyst Development

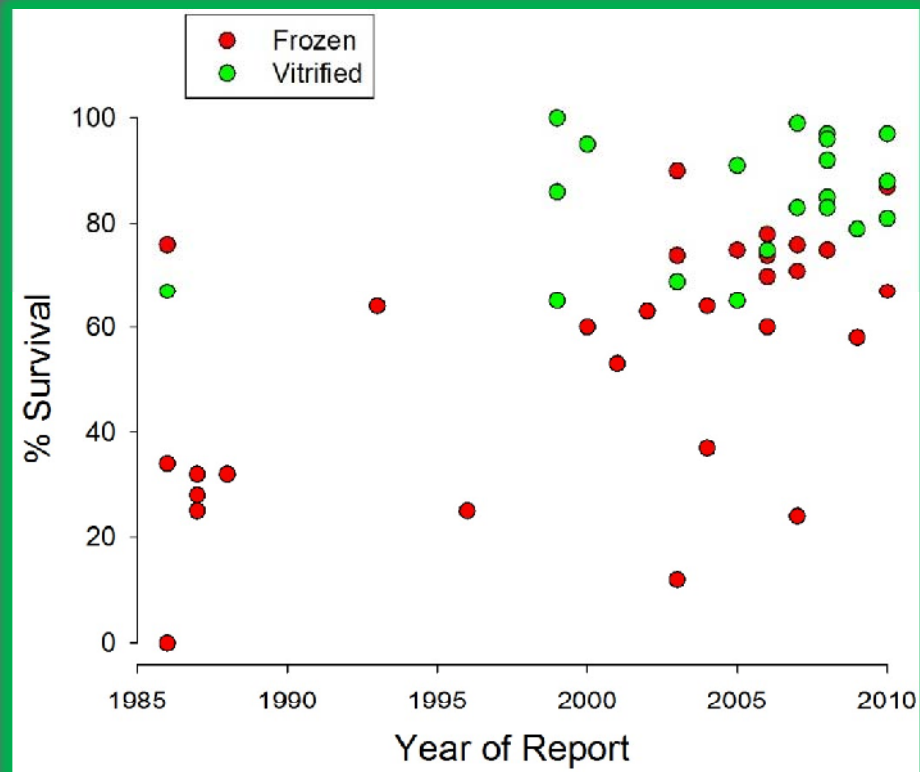
Control = 40%

5 seconds at 0 °C ≈ 20%



Martino A, Songsasen N, Leibo SP. Development into blastocysts of bovine oocytes cryopreserved by ultra-rapid cooling. *Biol Reprod* 1996;54:1059-69.

Human Oocyte Cryopreservation Outcomes



Today

Figure from Steven Mullen, The World Egg Bank, Phoenix AZ USA. See Reference List



Image from Irvine Scientific, Santa Ana, CA USA



Image from Kitazato, Shizuoka Japan

CURRENT VITRIFICATION METHODS

The Good, The Bad, and The Ugly

Technically Demanding

For skill embryologists/technicians:

SOP



1. Observation during the first week of training
2. Start training with spare material (IVM or unfertilized oocytes), under the guidance of the supervisor. Signed informed consent (the material will be used for teaching purposes).
3. Daily training during approximately 2 months (Estimated, depending on the skills and material available).
4. Start loading 1 oocyte per device and increase the number sequentially to be able to load 4 oocytes per device, in strict compliance with the protocol directions.
5. Over 80% SV in approximately 200-250 oocytes.
6. Beginners are not allowed to perform entire cases.



The challenge of vitrifying oocytes

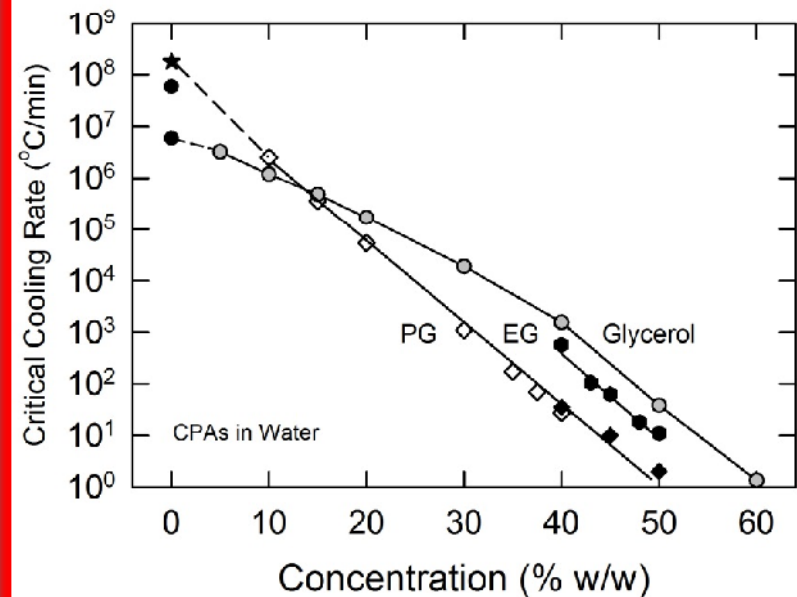
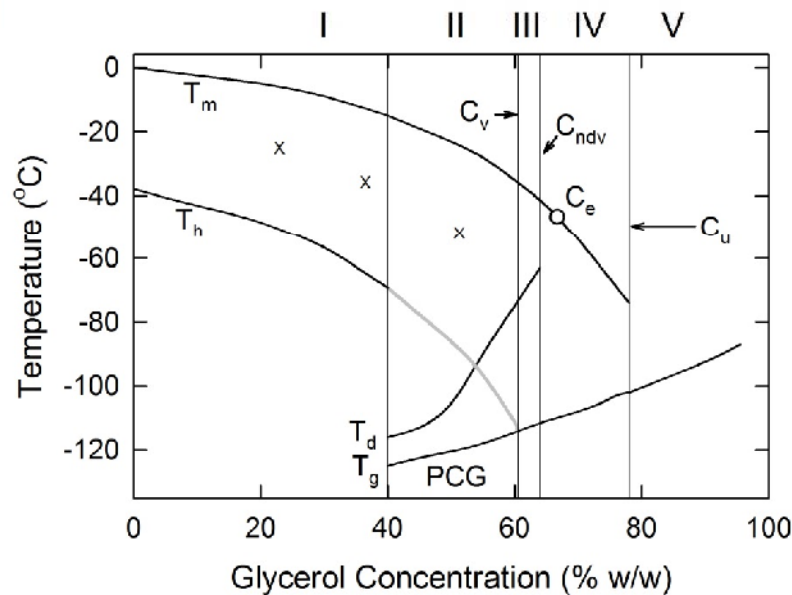


Image from Kitazato, Shizuoka Japan

Are We Really Vitrifying??

“...survival alone is not unequivocal proof of vitrification.”

Mullen SF, Fahy GM. Fundamental Aspects of Vitrification as a Method of Reproductive Cell, Tissue, and Organ Cryopreservation. In: Donnez J, Kim SS (eds), Principles and Practice of Fertility Preservation. Cambridge: Cambridge University Press, 2011;145-63.



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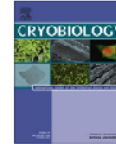
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Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions[☆]

Jesse B. Hopkins, Ryan Badeau, Matthew Warkentin, Robert E. Thorne^{*}

Physics Department, Cornell University, Ithaca, NY 14853, USA

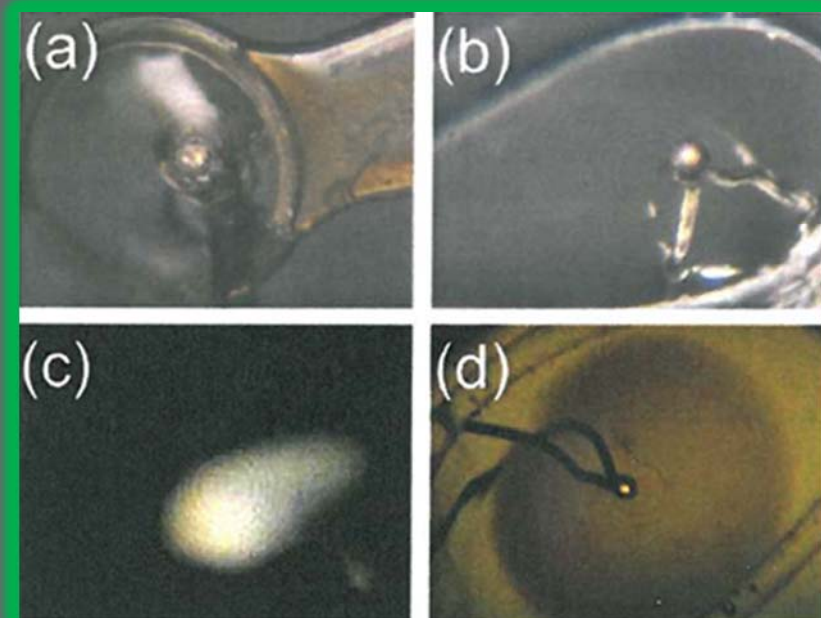
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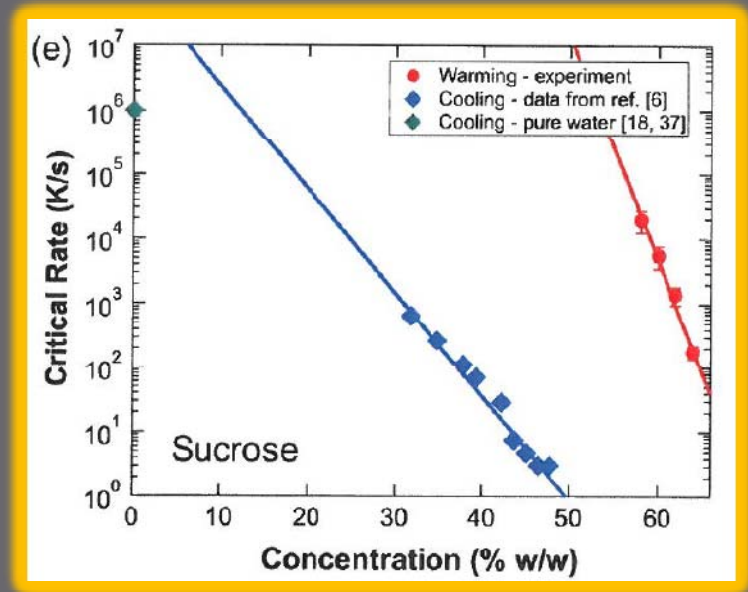
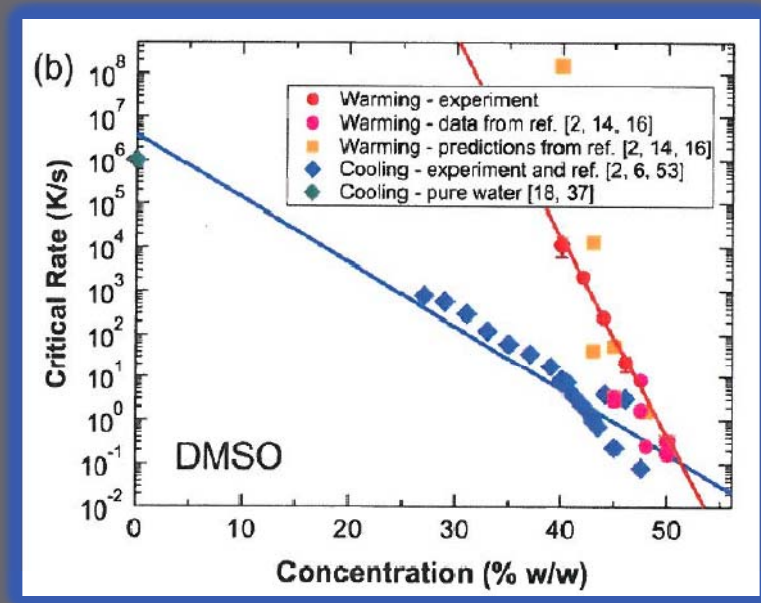
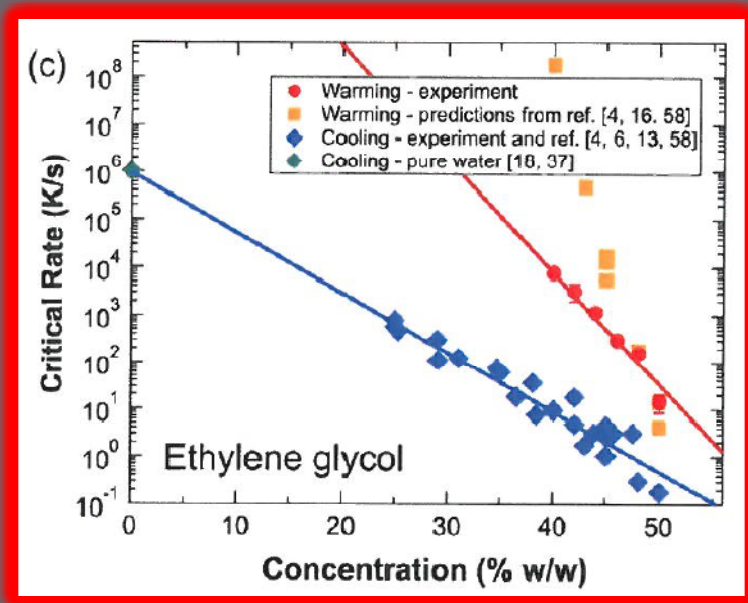
ABSTRACT

Ice formation on warming is of comparable or greater importance to ice formation on cooling in determining survival of cryopreserved samples. Critical warming rates required for ice-free warming of vitrified aqueous solutions of glycerol, dimethyl sulfoxide, ethylene glycol, polyethylene glycol 200 and sucrose have been measured for warming rates of order 10^{-10^4} K/s. Critical warming rates are typically

Finally!!

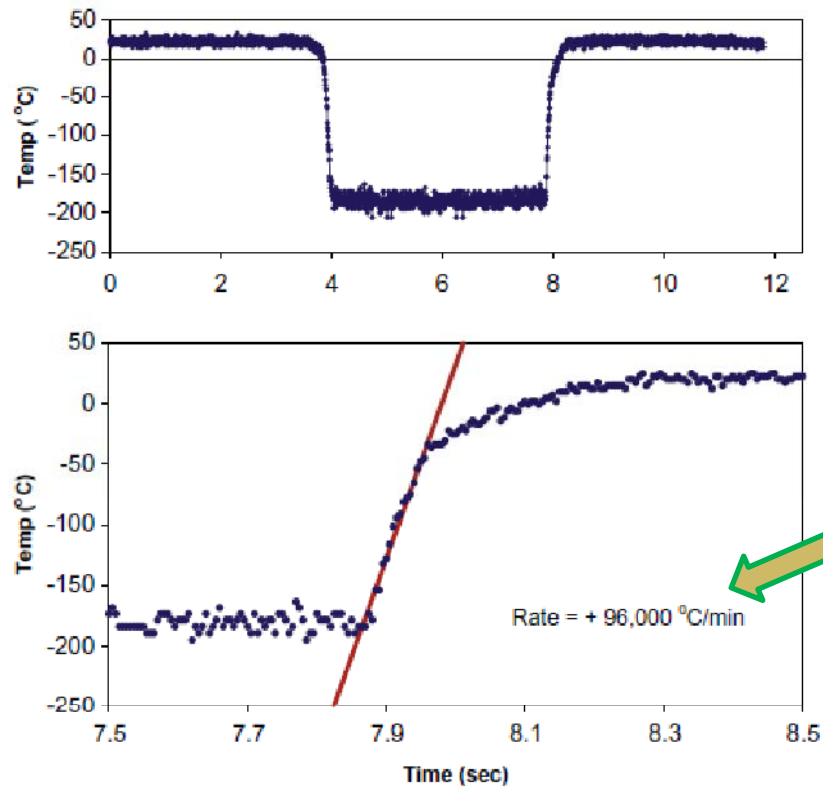


Hopkins JB, Badeau R, Warkentin M, Thorne RE. Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions. *Cryobiology* 2012;65:169-78.



Relevant Critical Warming Rates

Hopkins JB, Badeau R, Warkentin M, Thorne RE. Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions. *Cryobiology* 2012;65:169-78.

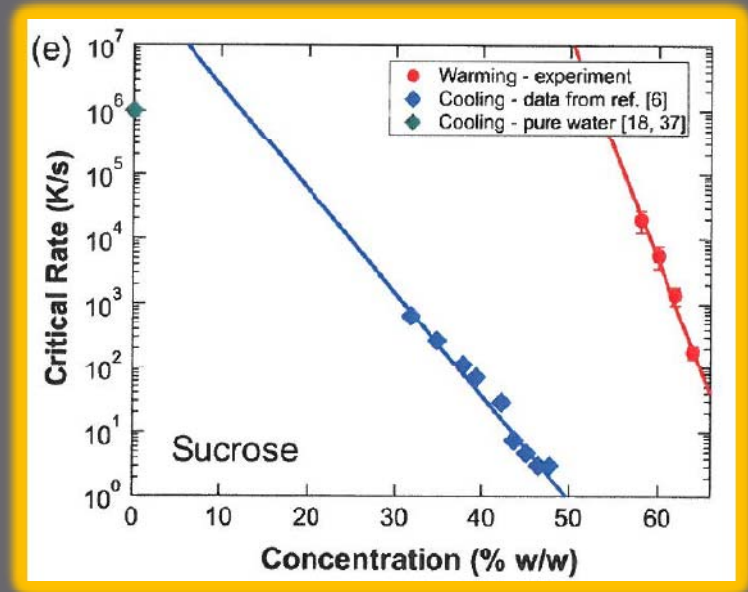
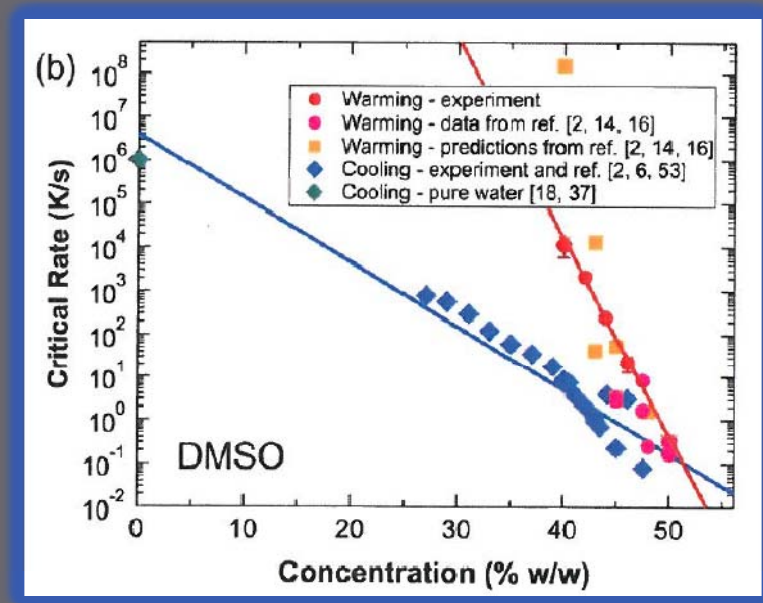
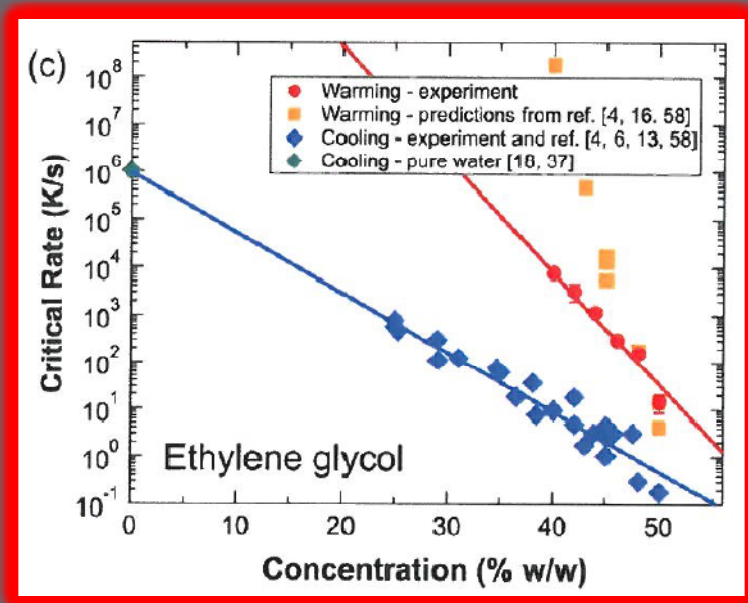


Warming Rate 96,000 K/min
 (= 1.6×10^3 K/sec)



Kleinhans FW, Seki S, Mazur P. Simple, inexpensive attainment and measurement of very high cooling and warming rates. *Cryobiology* 2010;61:231-3.

Image from Kitazato, Shizuoka Japan



**What about 15% DMSO
15% EG 0.5 mol/L Sucrose, Etc ????**
**Current Warming for Cryotop
about 10^3 K/s**

“Our preliminary attempts to measure critical cooling and warming rates in aqueous solutions containing up to 50% w/w lysozyme suggest that soluble proteins are very poor cryoprotectants, as has been found in studies of ice formation in hydrated protein powders.”



Hopkins JB, Badeau R, Warkentin M, Thorne RE. Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions. *Cryobiology* 2012;65:169-78.

Image from Kitazato, Shizuoka Japan

Big Unknown



Image from Kitazato, Shizuoka Japan

What is the vitrification solution composition in this scenario?

Shouldn't we Know??

Moving On...



Photo by S. F. Mullen

Thermal Sensitivity...

Seconds of Exposure to Room Temperature	Percent of Viable Embryos 5 hours post-warming
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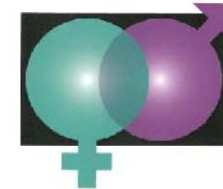
1	100
5	80
10	0
30	0
60	0



Image from Kitazato, Shizuoka Japan

P-40

Vitrified Embryos Are Vulnerable to Temperature Change. Charity Reeves, B.Sc., T.S.,^a Marlana Duke, M.S.,^a Lawrence Grunfeld, M.D.,^{a,b} Benjamin Sandler, M.D.,^{a,b} Tannoy Mukherjee, M.D.,^{a,b} Alan B. Copperman, M.D.^{a,b} ^aReproductive Medicine Associates of New York, 635 Madison Avenue, 10th Floor, New York, NY 10022; ^bDepartment of Obstetrics, Gynecology and Reproductive Science, Mount Sinai School of Medicine, Klingenstein Pavilion, 1176 Fifth Avenue, 9th Floor, New York, NY 10029.



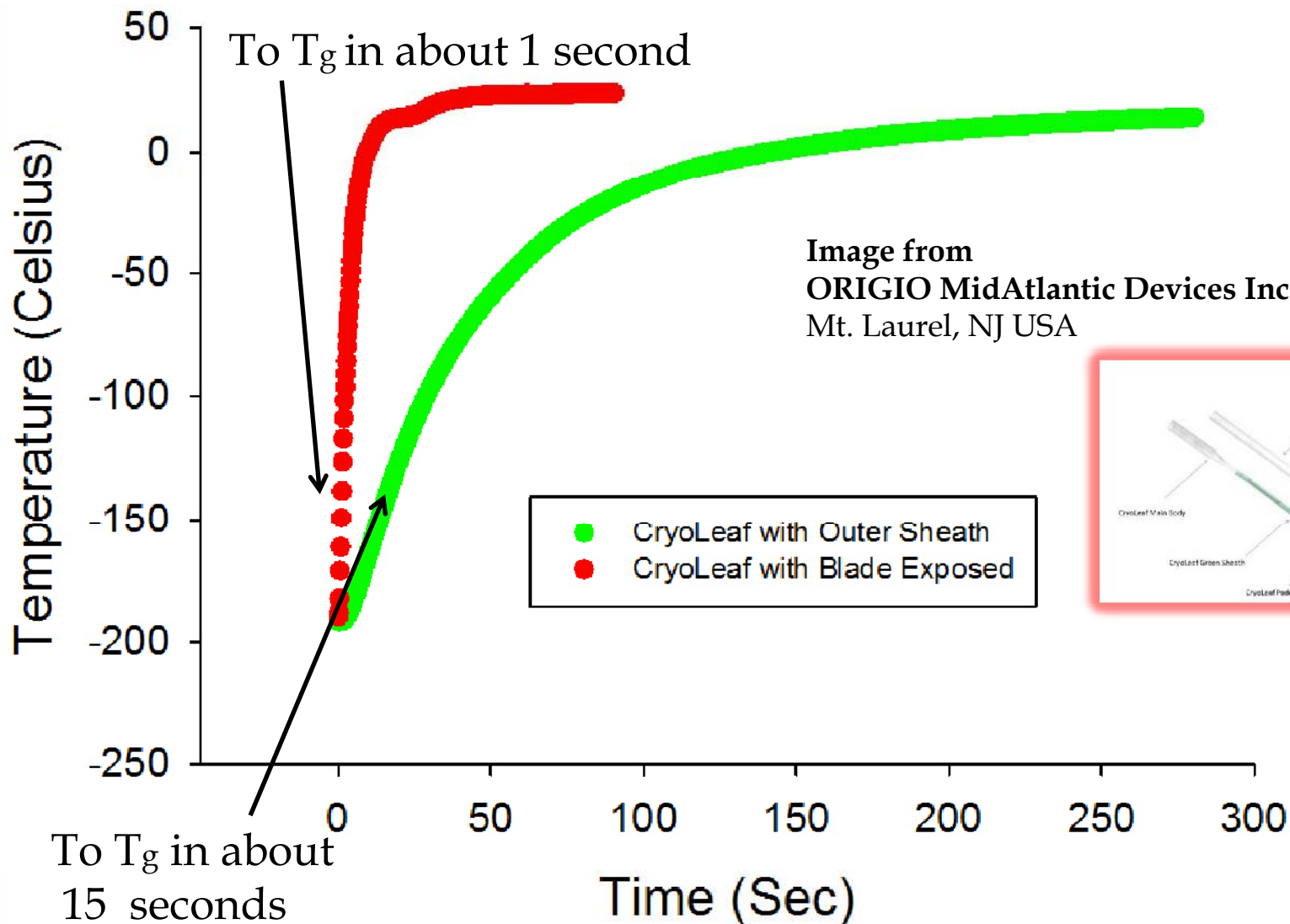
PACIFIC COAST
REPRODUCTIVE
SOCIETY

Abstracts of Oral and Poster Presentations

51st Annual Meeting of The
Pacific Coast Reproductive Society

The Future of Reproductive Medicine:
Creating a Health Care System

April 17 to April 21, 2013
Program Supplement



Data from Steven Mullen, The World Egg Bank, Phoenix AZ USA

So...what does this mean in practice ?

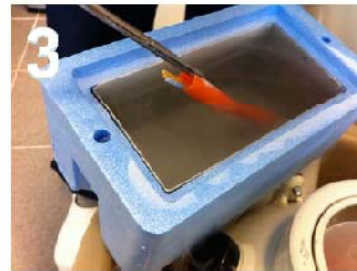
Sample reception at the destination



1 Fill the Dry Shipper with LN



2 Bring a recipient with LN



3 Take the gobelet to the container quickly



4 Storage



Temperature

The challenge of vitrifying cocytes

ALPHA 2012
Specialty in Reproductive Medicine
9th BIENNIAL CONFERENCE
APRIL 27 - 29, 2012, LONDON / UK
Church House - Conference Centre Westminster

The Challenge of vitrifying oocytes

Ana Cobo
IVI Valencia, Spain
ana.cobo@ivi.es
www.ivi.es

Moving On...

Mechanical Sensitivity...

Massive Fracture Event in Vitrified Solution in a Straw



Figure from Steven Mullen, The World Egg Bank,
Phoenix AZ USA



Movie from Rebecca De La Cruz and Steven Mullen, The World Egg Bank,
Phoenix AZ USA

What to Take Away...

- Vitrification research has a long history, and we are probably re-visiting many issues already investigated
- Vitrification, as is currently practiced, is technically demanding and requires a lot of practice and training to become proficient
- There are a lot of details that are simply unknown, and can lead to increased variability and difficulties in troubleshooting problems

These include:

- Whether vitrification is actually occurring
 - To what solution composition the cells are being exposed
 - How sensitive the samples are to accidental warming
 - How sensitive the samples are to thermomechanical stress
-
- Do we really need to go to these extremes?

References

- [1] Trounson A. Preservation of human eggs and embryos. *Fertil Steril* 1986;46:1-12.
- [2] Chen C. Pregnancy after human oocyte cryopreservation. *Lancet* 1986;1:884-6.
- [3] Al-Hasani S, Diedrich K, van der Ven H, Reinecke A, Hartje M, Krebs D. Cryopreservation of human oocytes. *Hum Reprod* 1987;2:695-700.
- [4] van Uem JF, Siebzehnruhl ER, Schuh B, Koch R, Trotnow S, Lang N. Birth after cryopreservation of unfertilized oocytes. *Lancet* 1987;1:752-3.
- [5] Diedrich K, Al-Hasani S, Van der Ven H, Krebs D. Successful in vitro fertilization of frozen-thawed rabbit and human oocytes. *Proceedings of the 5th world congress on IVF and ET* 1987:652-70.
- [6] Gook DA, Osborn SM, Johnston WI. Cryopreservation of mouse and human oocytes using 1,2-propanediol and the configuration of the meiotic spindle. *Hum Reprod* 1993;8:1101-9.
- [7] Tucker M, Wright G, Morton P, Shanguo L, Massey J, Kort H. Preliminary experience with human oocyte cryopreservation using 1,2-propanediol and sucrose. *Hum Reprod* 1996;11:1513-5.
- [8] Porcu E, Fabbri R, Damiano G, Giunchi S, Fratto R, Ciotti PM, Venturoli S, Flamigni C. Clinical experience and applications of oocyte cryopreservation. *Mol Cell Endocrinol* 2000;169:33-7.
- [9] Fabbri R, Porcu E, Marsella T, Rocchetta G, Venturoli S, Flamigni C. Human oocyte cryopreservation: new perspectives regarding oocyte survival. *Hum Reprod* 2001;16:411-6.

- [10] Quintans CJ, Donaldson MJ, Bertolino MV, Pasqualini RS. Birth of two babies using oocytes that were cryopreserved in a choline-based freezing medium. *Hum Reprod* 2002;17:3149-52.
- [11] Fosas N, Marina F, Torres PJ, Jove I, Martin P, Perez N, Arnedo N, Marina S. The births of five Spanish babies from cryopreserved donated oocytes. *Hum Reprod* 2003;18:1417-21.
- [12] Boldt J, Cline D, McLaughlin D. Human oocyte cryopreservation as an adjunct to IVF-embryo transfer cycles. *Hum Reprod* 2003;18:1250-5.
- [13] Chen SU, Lien YR, Chao K, Lu HF, Ho HN, Yang YS. Cryopreservation of mature human oocytes by vitrification with ethylene glycol in straws. *Fertil Steril* 2000;74:804-8.
- [14] Chen ZJ, Li M, Li Y, Zhao LX, Tang R, Sheng Y, Gao X, Chang CH, Feng HL. Effects of sucrose concentration on the developmental potential of human frozen-thawed oocytes at different stages of maturity. *Hum Reprod* 2004;19:2345-9.
- [15] Borini A, Bonu MA, Coticchio G, Bianchi V, Cattoli M, Flamigni C. Pregnancies and births after oocyte cryopreservation. *Fertil Steril* 2004;82:601-5.
- [16] Chen SU, Lien YR, Chen HF, Chang LJ, Tsai YY, Yang YS. Observational clinical follow-up of oocyte cryopreservation using a slow-freezing method with 1,2-propanediol plus sucrose followed by ICSI. *Hum Reprod* 2005;20:1975-80.
- [17] Boldt J, Tidswell N, Sayers A, Kilani R, Cline D. Human oocyte cryopreservation: 5-year experience with a sodium-depleted slow freezing method. *Reprod Biomed Online* 2006;13:96-100.
- [18] Borini A, Sciajno R, Bianchi V, Sereni E, Flamigni C, Coticchio G. Clinical outcome of oocyte cryopreservation after slow cooling with a protocol utilizing a high sucrose concentration. *Hum Reprod* 2006;21:512-7.

- [19] Levi Setti PE, Albani E, Novara PV, Cesana A, Morreale G. Cryopreservation of supernumerary oocytes in IVF/ICSI cycles. *Hum Reprod* 2006;21:370-5.
- [20] Bianchi V, Coticchio G, Distratis V, Di Giusto N, Flamigni C, Borini A. Differential sucrose concentration during dehydration (0.2 mol/l) and rehydration (0.3 mol/l) increases the implantation rate of frozen human oocytes. *Reprod Biomed Online* 2007;14:64-71.
- [21] Chamayou S, Alecci C, Ragolia C, Storaci G, Maglia E, Russo E, Guglielmino A. Comparison of in-vitro outcomes from cryopreserved oocytes and sibling fresh oocytes. *Reprod Biomed Online* 2006;12:730-6.
- [22] De Santis L, Cino I, Coticchio G, Fusi FM, Papaleo E, Rabbellotti E, Brigante C, Borini A, Ferrari A. Objective evaluation of the viability of cryopreserved oocytes. *Reprod Biomed Online* 2007;15:338-45.
- [23] De Santis L, Cino I, Rabbellotti E, Papaleo E, Calzi F, Fusi FM, Brigante C, Ferrari A. Oocyte cryopreservation: clinical outcome of slow-cooling protocols differing in sucrose concentration. *Reprod Biomed Online* 2007;14:57-63.
- [24] Parmegiani L, Cognigni GE, Bernardi S, Ciampaglia W, Infante F, Pocognoli P, de Fatis CT, Troilo E, Filicori M. Freezing within 2 h from oocyte retrieval increases the efficiency of human oocyte cryopreservation when using a slow freezing/rapid thawing protocol with high sucrose concentration. *Hum Reprod* 2008;23:1771-7.
- [25] Fadini R, Brambillasca F, Renzini MM, Merola M, Comi R, De Ponti E, Dal Canto MB. Human oocyte cryopreservation: comparison between slow and ultrarapid methods. *Reprod Biomed Online* 2009;19:171-80.
- [26] Cobo, A. The challenge of vitrifying oocytes. Presented at the 9th Biennial Conference ALPHA 2012 UK

- [27] Noyes N, Knopman J, Labella P, McCaffrey C, Clark-Williams M, Grifo J. Oocyte cryopreservation outcomes including pre-cryopreservation and post-thaw meiotic spindle evaluation following slow cooling and vitrification of human oocytes. *Fertil Steril* 2010;94:2078-82.
- [28] Smith GD, Serafini PC, Fioravanti J, Yadid I, Coslovsky M, Hassun P, Alegretti JR, Motta EL. Prospective randomized comparison of human oocyte cryopreservation with slow-rate freezing or vitrification. *Fertil Steril* 2010.
- [29] Antinori M, Licata E, Dani G, Cerusico F, Versaci C, Antinori S. Cryotop vitrification of human oocytes results in high survival rate and healthy deliveries. *Reprod Biomed Online* 2007;14:72-9.
- [30] Hong SW, Chung HM, Lim JM, Ko JJ, Yoon TK, Yee B, Cha KY. Improved human oocyte development after vitrification: a comparison of thawing methods. *Fertil Steril* 1999;72:142-6.
- [31] Kuleshova L, Gianaroli L, Magli C, Ferraretti A, Trounson A. Birth following vitrification of a small number of human oocytes: case report. *Hum Reprod* 1999;14:3077-9.
- [32] Kuwayama M, Vajta G, Kato O, Leibo SP. Highly efficient vitrification method for cryopreservation of human oocytes. *Reprod Biomed Online* 2005;11:300-8.
- [33] Lucena E, Bernal DP, Lucena C, Rojas A, Moran A, Lucena A. Successful ongoing pregnancies after vitrification of oocytes. *Fertil Steril* 2006;85:108-11.
- [34] Selman H, Angelini A, Barnocchi N, Brusco GF, Pacchiarotti A, Aragona C. Ongoing pregnancies after vitrification of human oocytes using a combined solution of ethylene glycol and dimethyl sulfoxide. *Fertil Steril* 2006;86:997-1000.

- [35] Yoon TK, Kim TJ, Park SE, Hong SW, Ko JJ, Chung HM, Cha KY. Live births after vitrification of oocytes in a stimulated in vitro fertilization-embryo transfer program. *Fertil Steril* 2003;79:1323-6.
- [36] Yoon TK, Lee DR, Cha SK, Chung HM, Lee WS, Cha KY. Survival rate of human oocytes and pregnancy outcome after vitrification using slush nitrogen in assisted reproductive technologies. *Fertil Steril* 2007;88:952-6.
- [37] Cao YX, Xing Q, Li L, Cong L, Zhang ZG, Wei ZL, Zhou P. Comparison of survival and embryonic development in human oocytes cryopreserved by slow-freezing and vitrification. *Fertil Steril* 2009;92:1306-11.
- [38] Cobo A, Kuwayama M, Perez S, Ruiz A, Pellicer A, Remohi J. Comparison of concomitant outcome achieved with fresh and cryopreserved donor oocytes vitrified by the Cryotop method. *Fertil Steril* 2008;89:1657-64.
- [39] Chang CC, Shapiro DB, Bernal DP, Wright G, Kort HI, Nagy ZP. Two successful pregnancies obtained following oocyte vitrification and embryo re-vitrification. *Reprod Biomed Online* 2008;16:346-9.
- [40] Sher G, Keskindepe L, Mukaida T, Keskindepe M, Ginsburg M, Agca Y, Maassarani G, Bayrak A. Selective vitrification of euploid oocytes markedly improves survival, fertilization and pregnancy-generating potential. *Reprod Biomed Online* 2008;17:524-9.
- [41] Chian RC, Huang JY, Tan SL, Lucena E, Saa A, Rojas A, Ruvalcaba Castellon LA, Garcia Amador MI, Montoya Sarmiento JE. Obstetric and perinatal outcome in 200 infants conceived from vitrified oocytes. *Reprod Biomed Online* 2008;16:608-10.
- [42] Kim TJ, Laufer LR, Hong SW. Vitrification of oocytes produces high pregnancy rates when carried out in fertile women. *Fertil Steril* 2010;93:467-74.

- [43] Rienzi L, Romano S, Albricci L, Maggiulli R, Capalbo A, Baroni E, Colamaria S, Sapienza F, Ubaldi F. Embryo development of fresh 'versus' vitrified metaphase II oocytes after ICSI: a prospective randomized sibling-oocyte study. *Hum Reprod* 2010;25:66-73.
- [44] Meryman HT. Basile J. Luyet: In Memoriam. *Cryobiology* 1975;12:283-92.
- [45] Hopkins JB, Badeau R, Warkentin M, Thorne RE. Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions. *Cryobiology* 2012;65:169-78.

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THANK YOU FOR
YOUR ATTENTION!